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Food anaphylaxis in antiphospholipid syndrome and thrombosis

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KEYWORDS

Antiphospolipid syndrome; thrombosis; food anaphylaxis; seeds; lipoproteins; oleosins

Abstract

Background: We have observed that some cases of food anaphylaxis were followed by severe thrombosis associated to anticardiolipin antibodies. Food anaphylaxis associated with antiphospholipid syndrome has seldom been published.

Objective: The aims were: 1) to test anticardiolipin antibodies in an important number of patients with anaphylaxis due to vegetal foods and their relationship with possible thrombosis; and 2) to study seed and fruit hypersensitivity in patients with previous thrombotic events associated with antiphospholipid antibodies (aCL).

Methods: We included 30 patients diagnosed of thrombosis associated with aCL, 52 patients who suffered from anaphylaxis due to seeds or fruits, and 120 control patients. Haematological, cardiopulmonary vascular and rheumatologic studies had been performed as needed. In vivo and in vitro allergy tests with a large battery of vegetal allergens were carried out in all the patients. Measurement of IgG aCL antibodies and specific IgE to vegetal food was done by ELISA and CAP-FEIA (Phadia). Immunodetection and inhibitions with lipoproteins belonging to seeds were performed.

Results: Seventy-five percent of the patients diagnosed as having antiphospholipid primary syndrome had specific IgE against different proteins from different vegetable allergens, most of them seeds, and clearly against lipoproteins that were also recognised by the patients with food anaphylaxis but not by the control cases. Among the patients with anaphylaxis, 28% had anticardiolipin antibodies and 17.3% thrombosis.

Conclusion: Our study suggests that seed lipoproteins which cause severe food anaphylaxis might have a potential role in the antiphospholipid syndrome and related thrombosis. © 2010 SEICAP. Published by Elsevier España, S.L. All rights reserved.

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Introduction

IgE-mediated reactions to fruits and vegetables are not uncommon in patients with allergic symptoms caused by pollen. Recently, different families of plant defence proteins, such as class I Chitinases, lipid transfer proteins (LTPs), cereal inhibitors of α -amylases and thaumatins have been identified as major allergens in allergies produced by plant foods and pollens. 2^{-4}

Severe food allergic reactions have been reported to involve the gastrointestinal, cutaneous, ocular, respiratory and cardiovascular systems. Anaphylactic reactions to foods almost always occur immediately and can be diagnosed by skin tests, specific IgE determination and food challenges if necessary, but the delayed clinical complications of this syndrome have not been completely determined yet.

We reported four patients with food-anaphylaxis followed by severe thrombosis. In all of them, anaphylaxis was due to the ingestion of vegetable allergens (seeds and fruits). The anaphylactic episode was followed by thrombosis of both iliac veins and lower cava. We found moderate-high levels of anticardiolipin (aCL) IgG antibodies and finally, the patients were diagnosed as having a primary antiphospholipid syndrome (APS) associated to anaphylaxis.

Here, we include 52 patients who suffered from anaphylaxis due to seeds and fruits in order to determinate levels of anticardiolipin (aCL) IgG antibodies and possible related symptoms. On the other hand, we have checked allergic hypersensitivity to seeds and fruits in 30 patients with previous thrombosis associated with aCL.

Methods

Patients

We reviewed the records of 21,879 patients who had been admitted to our Allergy Department in the last 22 years. We randomly selected 52 patients who suffered from severe anaphylaxis (grade III-IV by Müller) due to seeds or fruits. We also studied 30 patients with thrombosis and aCL antibodies, 20 having primary APS and 10 with APS secondary to systemic lupus erythematosus (SLE). As control group we included 120 subjects: 10 who suffered from anaphylaxis due to egg; 10 with anaphylaxis by Anisakis simplex; and 100 healthy non-atopic people. All the 202 subjects were studied in the same way: a detailed clinical allergy history followed by skin tests with a complete battery of 36 allergens, including prick-by-prick with suspected vegetal allergen, specific IgE measured by CAP method and lung function tests. Double blind placebo controlled oral challenge (DBPCOC) was performed only if necessary and in patients with non-severe anaphylaxis. Informed consent was obtained in all patients. Serum tests were done on individual samples from the 52 patients with anaphylaxis, 30 patients with thrombosis, as well as on pools from 100 non-allergic patients and from the 20 controls with anaphylaxis not due to vegetable foods.

Sensitisation to seeds or fruits was regarded as the presence of: a) one or more positive skin tests (with a wheal of 3 mm larger than the negative control to these allergens or

an area of $7\,\text{mm}^2$); b) a positive CAP-FEIA Phadia test >0.35 IU/mL; or c) a positive specific challenge. All these patients were informed of the objectives of the study and tested in order to try to identify an association of sensitisation to a vegetal allergen with thrombosis.

Seed and fruits extract

Commercial vegetable extract was supplied by ALK-ABELLO, Madrid, Spain (protein concentration 0.5 mg/mL). We performed extracts with fresh cereals seeds (wheat, barley, rye, and rice grains), legume seeds (soy bean, lentil, peanut, green pea), nuts (almond, hazelnut, chestnut, pine nut), garden produce (asparagus, tomato, leak), fruits (peach, apple, melon, banana), and mustard.

For in vitro tests, the different seeds and fruits were defatted, ground into small pieces and extracted by magnetic stirring in agitation in 50 mM phosphate-buffered saline (PBS) at pH 7.5 during 16 h at $4\,^{\circ}$ C. The samples were centrifuged at 5600 g for 30 minutes; the supernatant was dialysed against water. The dialysed extract was filtered through a 0.22 (m-pore diameter membrane and freezedried.

Skin prick tests

Skin prick tests (SPT) with extracts from different vegetables were performed according to standard procedures, over the anterior side of the forearm; one sterile device was used for each test. Histamine phosphate (10 mg/ml) and sterile 0.9% saline were used as positive and negative controls, respectively. A mean diameter of 3 mm or greater than the negative control and a mean area of 7 mm², 15 minutes after puncture, was considered a positive response. SPT with all these extracts were also performed in all patients of the control group.

Bronchial challenge tests

In patients in whom asthma was the predominant symptom of anaphylaxis, specific bronchial challenge tests (BCT) were done, following the procedure proposed by Chai et al., with modifications. Aerosolised particles generated by a continuous pressurised De Vilbiss 646 nebuliser were inhaled for 2 minutes at normal breathing volume, starting with control PBS solution, and followed - at 10 minutes intervals - by progressive concentrations of the extract. Pulmonary function tests were obtained 30 and 60 seconds after each dose. A positive response was defined as a greater than 20% fall from basal FEV1. After specific BCT, hourly peak expiratory flow (PEF) measurements were recorded for 9 hours. We used an extract of fresh-seed and fruits with a concentration 1/10 w/v.

Specific IgE measurements

Total IgE to different seed and fruits was determined by the CAP System IgE FEIA (Phadia Diagnostics, Uppsala, Sweden), following the manufacturer's instructions. The results were expressed in kU/L.

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Anticardiolipin antibodies

IgG anticardiolipin antibodies were measured by ELISA following the commercial recommendations (INOVA Diagnostics. Inc. Anticardiolipin IgG —HRP). Patients were tested again for anticardiolipin IgG antibodies after two months when positive. The laboratory criteria that we have adopted following the commercial manufacturer indications in the kit were: 10-20 GPL/ML low positive, 20-80 medium levels and >80 GPL/ML high positive.

SDS-PAGE Immunoblotting

We performed the immunodetection with hazelnut in selected patients sensitised to cereal grains, nuts and fruits. We selected hazelnut because it is the only allergen that can be used successfully in treatment with specific immunotherapy.⁹

The hazelnut liposoluble and hydrosoluble fractions were separated by 15% SDS-PAGE and transferred to PVDF membranes (Immobilon, MilliporeCo., USA). The transferred fractions were placed in contact with the patient sera, ⁶ and the IgE bound to the proteins was detected using mouse antihuman IgE (Fc)-HRP (SouthernBiotech, Birmingham, USA). Development in turn was based on chemiluminescence (ECL Plus WB Detection System, G.E. Healthcare, USA). (Fig. 1). Immunoblots were analysed in a GS-710 Image Analyser using the Diversity Database program (Bio Rad Laboratories, Hercules, CA, USA) following previously described methods.^{7–11}

ELISA inhibition

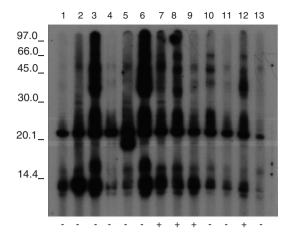
Plates were coated with 0.02 mg/mL of hydrosoluble proteins. Sera from patients who suffered from anaphylaxis were classified into two groups. Patients A had negative prick to seeds but positive IgE and immunodetection. Patients from group B presented positive prick, IgE and immunodetection. Patient sera from group A, patient sera from group B and both (A+B) were preincubated with hydrosoluble and liposoluble proteins at different protein concentration for two hours at room temperature. The bound IgE was detected by biotinylated mouse monoclonal antihuman IgE antibody (1:1000, 50 mcL/well; Operon, Zaragoza, Spain) followed by streptavidin-horseradish peroxidase-labelled antimouse IgG antibody (1:5000, 50 mcl/well; Sigma-Aldrich, St Louis, USA); IgE binding was detected by using a solution of 3,3',5,5'-tetramethylbenzidine (50 mcL/well; Sigma-Aldrich, St Louis, USA) and optical densities were read at 450 nm. (Fig. 2, 3).

Sera from patients not sensitised to fruit, as well as dilution buffer, were used as negative controls.

Statistical analysis

Data were analysed using the package SPPS for Windows v. 11.5 (SSPS Inc, 1989-1999 Chicago IL, EEUU).

Hydrosoluble Fr. Hazelnut



Liposoluble Fr. Hazelnut

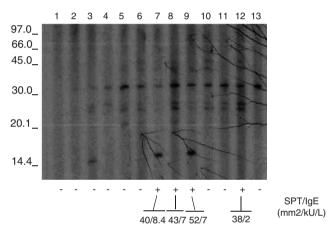


Figure 1 IgE Immunoblotting. Lanes 1-13: Patient's sera with anaphylaxis to nuts against hydrosoluble hazelnut fraction and liposoluble hazelnut fraction.

Informed consent

Written informed consent was obtained from all study participants.

Ethical Approval

Ethical approval was obtained from the Ethical Committee of the Rio Hortega University Hospital.

Results

The mean age of the patients who suffered from anaphylaxis was 20.9 ± 9.7 years. Of these patients, 21 were males and 31 females. Among the patients sensitised to nuts, all in group A (negative prick, positive IgE) were female, age 20 ± 6.2 years, whereas all in B were male, age 18 ± 2.6 years.

Among the 52 patients affected with anaphylaxis, 15 (28%) presented aCL (levels moderate-high) and related symptoms: 12 patients suffered from thrombosis: two of them of both iliac veins, four of either iliac or cava, one

1	Cer	eals Pric	k mm²/I	gE KU/L	L	egumes l	Prick/Ig	Ε		Nuts Pric	k/lgE		Vege	etables	Prick/IgE		Fruits p	rick/lgE		ACL Igo
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	36/4	32/3	20/2	19/1	-	-	-	-	-	19/1	-	-	-	-	-	23/22	-	-	-	10
	52/17	62/52	32/37	2072	22/2	-	-	-	24/3	-	-	-	-	-		32/3	22/1	-	-	2
	43/5	22/3	20/2	19/1	22/2	-	-	-	24/2	-	-	-	-	54/6		43/4	-	-	-	4
	32/2	45/6	25/2	-	20/2	-	-	-	-	-	-	-	-	-		42/5	-	-	-	2
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Anaphylaxis due to legumes: Patients 10 to 17: So: Soy bean, Le: lentil, Pe: peanut, Gp: Green pea. Anaphylaxis due to nuts: Patients 12 to 23: Al: Almond, Hn: Hazel nut, Cn: chestnut, Pn: Pine nut.		Ana	aphylaxi	is due to v	regetabl	es: Patients 24 to	29: As: As	paragus,	To. Tom	ato, Le:	leak.	·										
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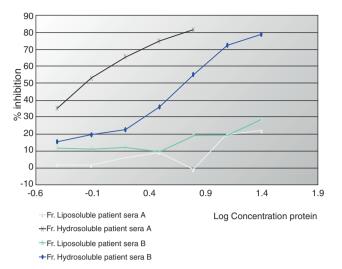


Figure 2 ELISA inhibition with "pools" of patients' sera with aCL that detected liposoluble fractions from seeds (oleosins). The patients that inhibited with oleosins (group A) were all women with primary APS.

Group A: Patients with Prick (-), IgE (+) and immunodetection (+).

Group B: Patients with Prick (+), lgE(+) and immunodetection (+).

intestinal bleeding due to mesenteric thrombosis and three women presented foetal losses. Their sensitisation to different allergens can be seen in Tables 1.

Among the 30 patients diagnosed previously of APS and thrombosis (Table 3), nine were male and 21 female, mean age 45.9 ± 19.9 years. Their mean age was significantly higher than the mean age of the persons with anaphylaxis (p < 0.0001). Both in the primary APS and in SLE, women were more common (60% and 90% respectively). As a whole - 15 patients- they all suffered from APS, and presented hypersensitivity to seeds and fruits, confirmed by allergy study. Two of these patients suffered anaphylaxis but they did not know the reason. None of the patients with SLE presented sensitisation to vegetables. None of the control patients had positive aCL levels. Symptoms and allergic response can be seen in Table 2.

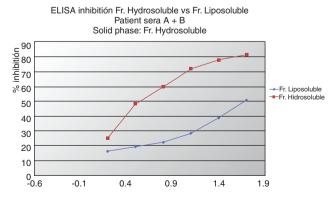


Figure 3 ELISA inhibition with "pools" of patients' sera with aCL that detected liposoluble fractions from seeds (oleosins). Groups A+B.

Table 1B Patients suffering from anaphylaxis by mustard and not by other seeds.

n	Must	ard Prick mm²/IgE KU/L	ACL: Ig	G (UGPL/mL
M1	56	7	9	
M2	39	4	12	
M3	68	8	6	
M4	29	3	8	
M5	62	19	61	
M6	66	4	7	
M7	78	45	5	
M8	27	6	4	
M9	30	6	3	
M10	52	9	12	

ACL: IgG (UGPL/mL).

Patients suffering from anaphylaxis by mustard.

Figures 1 and 2 shows the results of the immunoblotting and ELISA-inhibition experiments carried out. Several IgE-binding bands were detected both in hydrosoluble (ranging from around 25 to 100 KDa) and in liposoluble fractions (membrane lipoproteins) among the oleosins involved (with molecular masses around 16.7 and 14.7 kDa).

The recognition patterns obtained from the patients' sera presented a common result not found in any controls. The nine patients with negative prick showed negative specific IgE but a positive recognition of proteins from hydrosoluble and liposoluble fractions of hazelnut. All these patients were young women (20 ± 6 years). These responses were both soluble protein (up in the figure) and to lipoproteins (below). The response in patients with negative prick could be explained because the commercial available extracts are defatted, without lipoproteins.

In Figure 3 we can see an ELISA-inhibition test with sera pools from patients with aCL and which detected lipoproteins from seeds. The patient's sera that inhibited with lipoproteins were all from women suffering from APS.

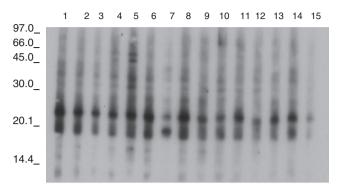
The sera from the 15 patients with primary APS showed IgE detection to lipoproteins belong to the membrane oleosins (16.7 and 14.7 KDa). (Figure 4). The membrane lipoproteins from the oleosin fraction were able to produce allergic sensitisation, a fact not demonstrated to date.

In summary, 75% of the patients diagnosed as having antiphospholipid primary syndrome had specific IgE against different proteins from different vegetable allergens, most of them seeds, and clearly against lipoproteins that were also recognised by the patients with food anaphylaxis but not by the control cases. Among the patients with anaphylaxis, 28% had anticardiolipin antibodies and 17.3% thrombosis.

Discussion

Anaphylaxis is a potential life-threatening condition. Prevention of anaphylaxis and anaphylactoid reactions concerns all physicians. To maximise the chances of preventing recurrences, the aetiology should be determined. 12 Nevertheless, it can be difficult to identify the causes, as well as to predict unexpected complications.

SAP patients Hazelnut Liposoluble fraction



Hazelnut Hydrosoluble fraction

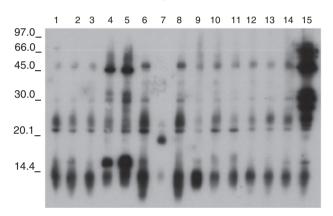


Figure 4 IgE-immunoblotting. Lanes 1 to 13. Patients' sera with primary APS against hydrosoluble hazelnut fraction and liposoluble hazelnut fraction.

This is the second known report of food anaphylaxis associated to primary antiphospholipid syndrome (APS). APS is diagnosed when arterial or venous thrombosis or recurrent miscarriages occur in a person in whom laboratory tests for antiphospholipid antibodies (aPL), namely lupus anticoagulant (LA), anticardiolipin antibodies (aCL) or antibodies directed to various proteins, mainly beta-2-glycoprotein (β_2 GPI), are positive.¹³

Evidence shows that some patients with antibodies to phospholipids are exposed to episodes of venous or arterial thrombosis, repeated foetal loss and thrombocytopenia. 14–17 It is also likely that the lupus anticoagulant and anticardiolipin antibody tests detect antibodies with overlapping specificities. 18 The term *primary* APS has been introduced as a mean of categorising and studying those patients with APS who do not meet criteria for SLE or any other well-defined diseases like infections or neoplasia. 17 Some clinicians have proposed that livedo reticularis, migraine headaches, chorea, peripheral artery occlusions, endocardial lesions, adult respiratory distress syndrome, avascular necrosis of the bone and transverse mielopathy might be features of APS. 17,18 Because of this overlap, the true clinical and serological boundaries of APS

1	Cer	eals Pricl	k mm²/lg	E KU/L	L	egumes l	Prick/lg	E		Nuts Pr	ick/lgE		Veg	getables P	rick/lgE		Fruits pr	rick/lgE		ACL IgG
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2	19/1	-	-	-	19/1	-	22/1		-	-	-	-	-	-	-	2271	-	-	-	48
3	28/3	46/3.3	24/2	36/3	38/2	32/1.5	34/3	20/1	20/1	32/1	24/2	-	-	28/2	-	48/2	20/1	30/2	42/2	48
4	66/15	96/54	32/32	34/3	22/1	32/1.2	42/2	-	-	64/12	32/12	-	46/3	46/3.4	-	-	32/1.2	42/0.9	46/2	76
5	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	32
6	47/2				25/1	-	-	-	26/2	26/1	-	-	-	-	-	20/1	-	-	-	98
7	-	-	-	-	2671	28/1	28/1	22/1	38/2	36/1	-	-	26/2	-	-	68/4	36/2	-	-	92
8	32/1	3071	32/1	36/1	30/1	38/1	36/1	26/1	-	28/1	-	-	52/2	36/1	-	42/2	38/3	-	-	106
9	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	105
0.	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	46
1	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	34/1	26/1	-	-	102
2	36/1	42/1	36/1	32/1	24/1	-	-	-	-	-	-	-	-	62/2	36/2	-	-	-	-	114
3	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	115
4	62/18	68/56	52/18	-	22/2	-	-	-	26/2	-	-	-	-	-	-	62/3	-	-	-	46
5	26/3	-	-	-	28/2	-	26/1	-	-	26/0.7	-	-	-	-	-	-	-	-	-	63
6	26/1	26/2	26/1	24/1	22/1	-	28/1	-	-	24/0.8	65/3	-	-	-	-	-	-	-	-	120
7	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	24
8	-	-	-	-	52/7	64/11	-	-	-	66/12	-	-	-	-	-	-	-	-	-	32
9	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	36
30	-	-	-	-	-	-	34/1	-	-	32/0.6	20/1	-	-	-	-	-	-	-	-	115

n	Age	Sex	Atopy	Clin Aller	Pathology
1	63	F	No	No	SLE
2	46	M	Si	No	APS, VT
3	57	F	No	No	APS, VT
4	43	F	No	No	SLE, VT
5	44	F	No	No	SLE, VT
6	51	F	No	No	SLE, VT
7	30	M	Yes	No	APS, VT
8	54	F	No	No	SLE
9	47	F	No	No	APS, VT
10	28	F	No	No	SLE
11	50	F	No	No	SLE
12	61	M	Yes	Anaphylaxis	APS, VT
13	83	M	No	No	APS, VT
14	54	F	No	No	APS, VT
15	73	M	No	No	APS,VT
16	43	F	Yes	No	APS IAM
17	51	F	No	Yes	APS,VT
18	10	M	Yes	Yes	APS, VT
19	43	F	No	No	SLE,
20	61	M	Yes	Yes	SLE
21	51	F	No	No	APS, VT
22	62	F	No	Yes	APS, VT
23	52	F	No	No	APS,VT
24	23	F	No	No	SLE, 1 abor
25	18	M	No	No	APS
26	64	F	No	No	APS, VT
27	32	F	No	No	APS, VT
28	34	F	Yes	Anaphylaxis	APS, 1 abor
29	27	F	No	No	APS,
30	22	M	No	No	APS, VT

are vague and will probably remain so until these patients are studied over time.

Given the heterogeneity of clinical manifestation in APS it is likely that more than one pathophysiological process may play a role.

In the case of our patients, aPL antibodies were persistent and could be related with the previous episodes of anaphylaxis. Deep vein thrombosis was the most frequent manifestation. In 3 cases, foetal losses were suffered recurrently.

The levels of aPL antibodies in our allergic patients were lower than those in the patients previously diagnosed of APS, but the age and evolution time in the last group was higher. In this respect aPL have been found in approximately 12% of elderly populations and in 2% of younger populations.¹³

Different genetic, environmental, $^{19-21}$ and immunological responses to infectious agents, 22 drugs or neoplasm 23 have been related with the production of aPL, but never with food allergy.

Food allergy hypersensitivity reactions have increased in the last decade, and are currently estimated to affect between 2–8% of the population, with a special incidence of food of plant origin in adult patients.²⁴ However, methods of diagnosis and treatment (potential immunotherapy), as well

as knowledge of the mechanisms of awareness, development of clinical symptoms and reactions to different allergenic sources (various foods and/or pollens), in general, are inadequate and incomplete.

Our patients' sera showed an IgE binding to several proteins included in hydrosoluble and liposoluble fractions (membrane lipoproteins) among which oleosins are involved. This response was not diagnosed under conventional protein extraction protocols. Then, membrane lipoproteins (oil body fraction contained) could have the capacity to sensitise.

The allergenic components of foods previously described include primarily glycoproteins. Despite the fact that closely related foods (legumes, cereal grains and other seeds) frequently contain allergens that cross-react immunologically (prick, RAST), they rarely cross-react clinically.²⁵ Allergy to *Rosaceae* fruits (peach, apple, pear) is frequently associated with birch pollinosis in The United States, Canada, and Central and Northern Europe. This cross-sensitisation has been explained mainly by the presence of homologous allergens in both the pollen from trees of the *Fagales* order and in the fruit/vegetables mentioned. Proteins of 16 to 18 Kd that share common IgE epitopes with Bet v 1, the major birch pollen allergen, have been characterised in alder, hazel and hornbeam pollens, and in celery, cherry and carrot. Nevertheless, profilin and Bet v 1-related structures

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are not involved in Rosaceae fruit allergy without pollinosis. In these patients the symptoms are usually more severe, as in our patients. ²⁶ The patients allergic to *Rosaceae* fruits and seeds whose sera were assayed in this study lived in cerealproducing areas free of birch and other Fagales trees and therefore could not be directly sensitised to their pollens. In this type of allergic population, which is frequent in the Mediterranean countries, a major 13 Kd allergen presented in various Rosaceae fruits has been detected. 1,2 In our study, the patients presented sensitisation to different vegetable foods, but Rosaceae fruits and seeds appear to have an important clinical role. Recently, we have purified and identified lipid transfer proteins (LTPs) from peach and wheat grain²⁷ and we have previously assayed in immunoblotting experiences a hazelnut extract that behaves as a very stable and useful product to investigate sensitisation to seeds and specific immunotherapy. We decided on the use of this extract to unify our results and to find something in common among the patients. Our results fully confirmed the relevance of glycoproteins and membrane lipoproteins allergens because they were recognised clearly by all the sera tested.

It was originally proposed that LTPs play a role in the cellular trafficking of lipids, but this hypothesis has now been largely abandoned because it is not consistent with several properties of this group of polypeptides, mainly with its extracellular location. By contrast, LTPs are truly involved in the defence mechanism of plants against pathogens. ²⁸ In this context, it is possible that the lipoproteins detected in this experience might be classified as defence proteins considering the uniformity and ubiquity of the antibodies in these patients.

Is it possible that there is a relationship between the sensitisation to these antigens and the APS suffered from our patients?

In this regard, it is tempting to suggest that these glyco and lipopoproteins, to which our patients had immunological responses, might have a similar behaviour to β_2 .GP-I, the necessary cofactor for the interaction between aCL and the antigen.²⁹ This is supported by the similar structure of human and vegetable membrane. Lipoproteins have a phospolipid bilayer shared by all eukaryotic cells⁸ including vascular endothelial and vegetal cells. Studies in APS have shown a remodelling of the membrane phospholipids bilayer in response to activation or apoptosis, that was defined as membrane-coated small vesicles that are membranecoated and released from the plasma membrane by exocitic budding. These vesicles express negatively charged phospholipids and cell surface antigens characteristics of the cells of origin.8 Surface exposure of phosphatidylserine or tissue factor activity provides a catalytic surface that supports the assembly of clotting enzymes complexes, leading to thrombin generation.

The thrombosis may be due to an alteration of the kinetics of the normal procoagulant and anticoagulant reactions by cross-linking membrane-bound proteins, by blocking protein-protein interactions or by blocking the access of other proteins to the phospholipid membrane.³⁰

There is a platelet role in anaphylaxis, and some data suggest that platelet activation in APS patients is related to anti β_2 -GP-I antibodies. ^{31,32} In this way, endothelial activation in APS has been described to be mediated through Toll-like receptors (TLR4), resulting in a prothrombotic and

pro-inflammatory phenotype with synthesis and secretion of adhesion molecules and pro-inflammatory cytokines very similar to those released during anaphylactic response to vegetal foods.³¹

The prevalence of food allergies has increased over the past fifteen years. Recent studies have found that 17% of young adults reported that particular food or foods nearly always cause illness or trouble when eaten, presumably due to either food intolerance or food allergy.³² The reasons suggested are changes in dietary behaviour and the evolution of food technologies. New food-processing techniques can increase the allergenicity or create neo-allergens; storage can also induce the synthesis of allergenic stress or PR proteins.³³ Genetically-modified plants have risks of allergenicity, and methodological steps of investigations as well as the studies with sera from allergic patients are required. On the other hand, the combination of enzymatic hydrolysis, heating, or the development of genetically-modified plants may offer new alternatives towards hypoallergenic foods.34

With respect to the treatment of APS with thrombosis, there is consensus in treating patients with oral anticoagulation to a target International Normalised Ratio of 2.0 to 3.0.³⁵ The approach for women with obstetric manifestations is based on the use of aspirin plus heparin.³⁶ In addition to this we recommended these measures and a diet without the putative vegetal allergen to our patients. None of our patients has suffered from new episodes of thrombosis in five years and one woman had her first child successfully.

In summary, despite the strong association between aPL and thrombosis, the pathogenic role of these antibodies in the development of thrombosis has not been fully elucidated. A novel mechanism involving hypersensitivity to lipoproteins allergens is described here. Nevertheless, our preliminary observations must be confirmed in further studies. The knowledge of these new pathogenic approaches might identify novel therapeutic targets and therefore may improve the management of these patients.

Conflict of interest

The authors report no conflict of interest.

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References

- Ortolani C, Pastorello EA, Farioli L, Ispano M, Pravettoni V, Berti C, et al. IgE mediated allergy from vegetable allergens. Ann Allergy. 1993;71:470-6.
- Diaz Perales A, Collada C, Blanco C, Sánchez-Monge R, Carrillo T, Aragoncillo C, et al. Class I chitinases with hevein-like domain, but not class II enzymes, are relevant chestnut and avocado allergens. J Allergy Clin Immunol. 1998;102:127–33.

- Sanchez-Monge R, Lombardero M, García Sellés FJ, Barber D, Salcedo G. Lipid transfer proteins are relevant allergens in fruit allergy. J Allergy Clin Immunol. 1999;103:514–9.
- Palacín A, Quirce S, Armentia A, Fernandez-Nieto M, Pacios LF, Sastre J, et al. Wheat lipid transfer protein is a major allergen associated with baker's asthma. J Allergy Clin Immunol. 2007 Nov:120:1132—8.
- 5. Armentia A, Barber D, Lombardero M, Martín-Santos JM, Martín Gil FJ, Arranz ML, et al. Anaphylaxis associated with antiphosphlipid syndrome. Ann Allergy. 2001;87:54—9.
- Towbin H, Staehelin T, Gordon J. Electrophoretic transfer of proteins from polyacrylamide gels to nitro-cellulose sheets; procedure and some applications. Proc Natl Acad Sci USA. 1974;76:4350–4.
- 7. Sánchez Madrid F, Morago G, Corbí AL, Carreira J. Monoclonal antibodies to three distinct epitopes on human IgE: their use for determination of allergen-specific IgE. J Immunol Methods. 1984;73:367—78.
- 8. Distler JH, Huber LC, Gay S, Sistler O, Pisetky DS. Microparticles as mediators of cellular cross-talk in inflammatory disease. Autoinmunity. 2006;39:638–90.
- Enrique F, Pineda F, Malet T, Bartra J, Balagaña M, Tella R, et al. Sublingual immunotherapy for hazelnut food allergy: a randomized, double-blind, placebo-controlled study with a standardized hazelnut extract. Journal of Allergy and Clinical Immunology. 2005;116:1073—9 (November 2005).
- 10. Jaap HA, Frauke S, Stefan V, Serge V, Laurian Z, Sue LH, et al. Cloning of oleosin, a putative new hazelnut allergen, using a hazelnut cDNA library. Mol. Nutr. Food Res. 2006, 50, 18–23.
- Armentia A, Sánchez Monge R, Gómez L, Barber D, Salcedo G. In vivo allergenic activities of eleven purified members of a major allergen family from wheat and barley flour. Clin Exp Allergy. 1993;23:410–5.
- 12. Nickolas RA, Bernstein IL, Li James T, Lee Rufus E, Spector SL, Dykewicz MS, et al. The diagnosis and management of anaphylaxis. J Allergy Clin Immunol. 1999;6:S465—98.
- 13. Espinosa G, Cervera R. Antiphospholipid síndrome. Arthritis Research & Therapy 2008; 10:230—239. Available online http://artrhitis-research.com/content/10/6/230.
- 14. Miyakis S, Lockshin MD, Atsumi T, Branch DW, Brey RL, Cervera R, et al. International consensus statement on an update of the classification criteria for definite antiphospholipid syndrome (APS). J Throm Haemos. 2006;4:295–306.
- 15. Vermylen J, Blockmans D, Spitz B, Deckmyn H. Thrombosis and immune disorders. Clin Haematol. 1986;15:393.
- Harris EN, Khamashta MA, Hughes GRV. Antiphospholipid antibody syndrome. In Arthritis and allied conditions. McCarty and Koopman Eds. Philadelphia, London 1993.
- 17. Hughes GRV. The anticardiolipin syndrome. Clin Exp Rheumatol. 1985:285—6.
- 18. Asheerson RA, Cervera R. Unusual manifestations of the antiphospholipid syndrome. Clin Rev Allergy Immunol. 2003;25:61–78.
- 19. Uthman I, Khamastha M. Ethnic and geographical variation in antiphospholipid (Hughes) syndrome. Ann Rhem Dis. 2005;64:1671-6.
- Yasuda S, Atsumi T, Matsuura E, Kaihara K, Yamamoto D, Ichikawa K, et al. Significance of valine/leucine247 polymorphism of beta2-glycoprotein I in antiphospholipid syndrome:

- increased reactivity of anti-beta2-glycoprotein I antibodies to the valine247 beta2-glycoprotein I variant. Arthritis Rheum. 2005:52:212-8.
- 21. Camilleri RS, Mackie IJ, Humphries SE, Machn Sj, Cohen H. Lack of association of beta2-glycoprotien polymorphisms Val1247Leu and Trp216Ser with antiphospholipid antibodies in patients with thrombosis and pregnancy complications. Br J Haematol. 2003;120:1066–72.
- 22. Blank M, Krause I, Fridkin M, Keller N, Kopolovic J, Goldberg I, et al. Bacterial induction of autoantibodies to beta 2 glycoprotein-I accounts for the infectious etiology of beta2-glycoprotein accounts for the infectious etiology of antiphospholipid syndrome. J Clin. 2002;109:797–804.
- 23. Miesbach W, Asheron RA, Cervera R, Shoenfeld Y, Gomez Puerta J, Bucciarelli S, et al., Members of CAPS Registry Group. The catastrophic antiphospholipid (Asherson's) syndrome and malignancies. Autoinmun Rev. 2006;6:94–7.
- 24. Sampson HA. Food allergy. JAMA. 1997;278:1888-94.
- 25. Pastorello EA, Farioli L, Pravettoni V, Ortolani C, Ispano M, Monza M, et al. The major allergen of peach (Prunus persica) is a lipid transfer protein. J Allergy Clin Immunol. 1999;103:520—6.
- Fernández Rivas M, van Ree R, Cuevas M. Allergy to Rosaceae fruits without related pollinosis. J Allergy Clin Immunol. 1997;100:728–33.
- Palacín A, Quince S, Armentia A, Fernández-Nieto M, Pacios LF, Asensio T, et al. Wheat lipid transfer protein is a major allergen associated with baker's asthma. J Allergy Clin Immunol. 2007;120:1132–8.
- 28. Salcedo G, Diaz-Perales A, Sanchez Monge R. Fruit Allergy: plant defence proteins as novel potential panallergens. Clin Exp Allergy. 1999;29:1158—60.
- 29. Cortés T. Síndrome antifosfolipidos. Diagnostico de laboratorio. Ed Cont Lab Clin. 1999;2:25—32.
- 30. Shi W, Chong BH, Chesterman CN. β2 glycoprotein I is a requirement for anticardiolipin antibodies binding to activates platelets: differences with lupus anticoagulant. Blood. 1993;81:1255–62.
- Pierangeli SS, Vega-Ostertag ME, Raschi E, Liu X, Romay-Penabad Z, DE Micheli V, et al. Toll-like receptor and antiphsopolipid mediated thrombosis: in vivo studies. Ann Rheum Dis 2007; 66:1327–1333.
- 32. Woods RK, Abramson M, Raven JM, Bailey M, Weiner JM, Walters EH. Reported food intolerance and respiratory symptoms in young adults. Eur Respir J. 1998;11:151–5.
- 33. Moneret-Vautrin DA. Modifications of allergenicity linked to food technologies. Allerg Immunol. 1998;30:9—13.
- 34. de Gregorio M, Armentia A, Díaz-Perales A, Palacín A, Dueñas-Laita A, Martín B, Salcedo G, et al. Salt-soluble proteins from wheat-derived foodstuffs show lower allergenic potency than those from raw flour. J Agric Food Chem. 2009 Apr 22;57:3325—30.
- 35. Ruiz-Irastorza G, Hunt BJ, Khamashta MA. A systematic review of secondary thromboprophylaxis in patients with antiphospholipid antibodies. Arthritis Rheum. 2007;57: 1487–95.
- Derksen RHm, Khamashta MA, Branch DW. Management of the obstetric antiphospholipid syndrome. Arthritis Rheum. 2004;50:1028–39.