

Short communication

[Dmt¹, D-1-Nal³]morphiceptin, a novel opioid peptide analog with high analgesic activity

Jakub Fichna^a, Jean-Claude do-Rego^b, Nga N. Chung^c, Jean Costentin^b, Peter W. Schiller^c, Anna Janecka^{a,*}

^a Laboratory of Biomolecular Chemistry, Institute of Biomedicinal Chemistry, Medical University of Lodz,

Mazowiecka 6/8, 92-215 Lodz, Poland

^b Laboratoire de Neuropsychopharmacologie Expérimentale, CNRS-FRE 2735, IFRMP 23, Université de Rouen, France

^cLaboratory of Chemical Biology and Peptide Research, Clinical Research Institute of Montreal, Montreal, Canada

ARTICLE INFO

Article history: Received 4 November 2007 Received in revised form 7 December 2007 Accepted 7 December 2007 Published on line 23 December 2007

ABSTRACT

The morphiceptin-derived peptide [Dmt¹, p-1-Nal³]morphiceptin, labeled μ -opioid receptor (MOP) with very high affinity and selectivity in the receptor binding assays. In the mouse hot plate test, [Dmt¹, p-1-Nal³]morphiceptin given intracerebroventricularly (i.c.v.) produced profound supraspinal analgesia, being approximately 100-fold more potent than the endogenous MOP receptor ligand, endomorphin-2. The antinociceptive effect of this new analog lasted up to 120 min. Thus, [Dmt¹, p-1-Nal³]morphiceptin is an interesting and extraordinarily potent analgesic, raising the possibility of novel approaches in the design of clinically useful drugs for pain treatment.

© 2007 Elsevier Inc. All rights reserved.

1. Introduction

Endomorphin-1 (Tyr-Pro-Trp-Phe-NH₂, EM-1) and endomorphin-2 (Tyr-Pro-Phe-Phe-NH₂, EM-2) are two endogenous opioid peptides isolated from the mammalian brain which show remarkably high selectivity for the μ -opioid receptor (MOP receptor) [28]. Endomorphins exhibit pharmacological properties similar to those of morphine and also produce morphine-like undesirable side effects upon exogenous administration (for review see [10]).

In efforts to develop new candidate drugs with antinociceptive activity, numerous chemical modifications have been performed in order to improve the pharmacological profile of endomorphins [3,13,16,17,27,29]. Structurally related to EM-2 is morphiceptin (Tyr-Pro-Phe-Pro-NH₂), which does not occur in neuronal tissue but, like EM-2,

displays morphine-like physiological activity and is very selective for the MOP receptor [4]. The two aromatic amino acids in EM-2 and morphiceptin, Tyr¹ and Phe³, are important structural elements in the interaction with the receptor. Hansen et al. first showed that replacing Tyr¹ in the cyclic opioid peptide H-Tyr-c[D-Pen-Gly-Phe-D-Pen]NH₂ (DPDPE) with the more hydrophobic 2',6'-dimethyltyrosine (Dmt) residue resulted in a compound with greatly increased MOP and DOP receptor binding affinities and much improved antinociceptive potency [12]. Subsequently, it was reported that substitution of Dmt for Tyr¹ in various MOP receptor agonist peptides or mixed MOP receptor agonist/ DOP receptor antagonist peptides also greatly enhanced MOP receptor agonist potency and antinociceptive activity [2,5,11,19,22–26]. Furthermore, we showed that introduction of a D-1-naphthylalanine (D-1-Nal) residue in place of Phe³ in

^{*} Corresponding author. Tel.: +48 42 679 04 50x259; fax: +48 42 678 42 77. E-mail address: ajanecka@zdn.am.lodz.pl (A. Janecka).

^{0196-9781/\$ –} see front matter © 2007 Elsevier Inc. All rights reserved. doi:10.1016/j.peptides.2007.12.005

morphiceptin increased MOP receptor binding affinity and associated functional bioactivity 40- and 30-fold, respectively, and elevated the *in vivo* activity in mice, as measured by the classical hot plate test of analgesia [6].

In the present study, [D-1-Nal³]analogs of morphiceptin and EM-2 were synthesized and further modified by substitution of a Dmt residue in position 1 in place of Tyr. The pharmacological profiles of the new analogs were determined in *in vitro* and *in vivo* assays.

2. Materials and methods

2.1. Materials

Morphiceptin, EM-2, and their analogs were synthesized by standard solid phase procedures as described before [8]. [³H]DAMGO and [³H]naltrindole were purchased from PerkinElmer-NEN Life Science Products (Paris, France).

2.2. Methods

2.2.1. Opioid receptor binding assays

The MOP and DOP receptor binding studies were performed according to the modified method described earlier [9]. Crude membrane preparations, isolated from Wistar rat brains, were incubated at 25 °C for 120 min with appropriate concentration of a tested peptide in the presence of either 0.5 nM [³H]DAMGO or 0.5 nM [³H]naltrindole in a total volume of 0.5 ml of 50 mM Tris-HCl (pH 7.4), containing MgCl₂ (5 mM), BSA (1 mg/ml), bacitracin (50 µg/ml), bestatin (30 μ g/ml), and captopril (10 μ M). Non-specific binding was determined in the presence of 10 µM naloxone. Incubations were terminated by rapid filtration through Whatman GF/B (Brentford, UK) glass fiber strips, which had been presoaked for 2 h in 0.5% polyethylamine, using Sampling Manifold (Millipore, Billerica, MA, USA). The filters were washed three times with 4 ml of ice-cold Tris buffer solution. The bound radioactivity was measured in a Tri-Carb 2100 TR liquid scintillation counter (Packard, Ramsey, MN, USA) after overnight extraction of the filters in 4 ml of Ultima Gold scintillation fluid (PerkinElmer, Wellesley, MA, USA). Three independent experiments for each assay were carried out in duplicate.

2.2.2. In vitro bioassays

The GPI and MVD assays were performed as described previously [23].

2.2.3. Antinociception studies

The hot plate test in mice was performed as described previously [6], according to the European Communities Council Directive from 24 November 1986 (86/609/EEC) and were conducted by authorized investigators.

Table 1 – Physicochemical data of morphiceptin and endomorphin-2 analogs						
No.	Sequence	HPLC t _r ^a (min)	FAB-MS			Purity (%)
			Formula	MW	$[M + H]^{+}$	
1	Tyr-Pro-Phe-Pro-NH ₂ (morphiceptin)	8.15	C ₂₈ H ₃₅ N ₅ O ₅	521	522	98
2	Tyr-Pro-d-1-Nal-Pro-NH2	11.36	C ₃₂ H ₃₇ N ₅ O ₅	571	572	97
3	Dmt-Pro-d-1-Nal-Pro-NH2	14.86	$C_{34}H_{41}N_5O_5$	599	600	97
4	Tyr-Pro-Phe-Phe-NH ₂ (endomorphin-2)	12.51	C ₃₂ H ₃₇ N ₅ O ₅	571	572	98
5	Tyr-Pro-d-1-Nal-Phe-NH2	16.05	$C_{36}H_{39}N_5O_5$	621	622	98
6	$Dmt-Pro-d-1-Nal-Phe-NH_2$	17.06	$C_{38}H_{43}N_5O_5$	649	650	97

^a HPLC elution on a Vydac C_{18} column (0.46 cm \times 25 cm, 5 μ m) using the solvent system of 0.1% TFA in water (A)/80% acetonitrile in water containing 0.1% TFA (B) and a linear gradient of 20–80% solvent B over 25 min at flow rate of 1 ml/min.

Table 2 – Competitive binding and functional assay data obtained with morphiceptin and endomorphin-2 analogs						
No.	Sequence	IC ₅₀ (nM)				
		MOP ^a	DOP ^b	DOP/MOP	GPI	MVD
1	Tyr-Pro-Phe-Pro-NH ₂ (morphiceptin)	19.8 ± 0.9	>1000	>50.5	$318\pm\mathbf{22^{c}}$	4800 ± 440^{c}
2	Tyr-Pro-d-1-Nal-Pro-NH ₂	$\textbf{0.47}\pm\textbf{0.05}$	>1000	>2127	$9.57 \pm 1.01^{\text{c}}$	$\textbf{35.4} \pm \textbf{2.3}^{c}$
3	Dmt-Pro-D-1-Nal-Pro-NH ₂	0.000549 ± 0.000145	132 ± 31	>13000	$\textbf{0.45} \pm \textbf{0.06}$	$\textbf{0.64} \pm \textbf{0.06}$
4	Tyr-Pro-Phe-Phe-NH ₂ (endomorphin-2)	$\textbf{0.79} \pm \textbf{0.05}$	>1000	>1265	$\textbf{7.71} \pm \textbf{1.47}$	15.3 ± 1.8
5	Tyr-Pro-D-1-Nal-Phe-NH ₂	$\textbf{22.3}\pm\textbf{0.9}$	>1000	>44.8	872 ± 90	2040 ± 290
6	Dmt-Pro-d-1-Nal-Phe-NH ₂	$\textbf{0.93}\pm\textbf{0.13}$	>1000	>1075	$\textbf{1.53} \pm \textbf{0.27}$	$\textbf{1.63} \pm \textbf{0.09}$
	Tyr-D-Ala-Phe-Glu-Val-Val-Gly-NH ₂ (deltorphin-II)	-	$\textbf{0.56} \pm \textbf{0.05}$	-	-	-
	[Leu ⁵]enkephalin	-	-	-	246 ± 39	11.4 ± 1.1

All values are expressed as mean \pm S.E.M. of three to six determinations.

^a Determined against [³H]DAMGO.

^b Determined against [³H]naltrindole.

^c Data from Ref. [6].



Fig. 1 – Dose–response curves determined in the hot plate test for the inhibition of paw licking (A), rearing (B), and jumping (C) induced by i.c.v. injection of morphiceptin, EM-2, $[Dmt^1, D-1-Nal^3]$ morphiceptin (analog 3), and $[Dmt^1, D-1-Nal^3]$ EM-2 (analog 6). The data represent the mean \pm S.E.M. of 10 mice per group.



Fig. 2 – Time-course of the changes in inhibition of paw licking (A), rearing (B), and jumping (C) induced by i.c.v. injection of morphiceptin (10 μ g), EM-2 (3 μ g), and [Dmt¹, D-1-Nal³]morphiceptin (analog 3, 0.1 μ g), determined in the hot plate assay.

injection (10 μg/animal)						
No.	Sequence	L	Latencies (%MPE) to			
		Paw licking	Rearing	Jumping		
1	Tyr-Pro-Phe-Pro-NH $_2$ (morphiceptin) ^a	11.52 ± 4.19	$\textbf{33.08} \pm \textbf{6.54}$	$\textbf{75.12} \pm \textbf{8.48}$	1.75	
2	Tyr-Pro-D-1-Nal-Pro-NH2 ^a	44.48 ± 8.63	$\textbf{75.84} \pm \textbf{6.63}$	92.72 ± 5.57	0.50	
3	Dmt-Pro-d-1-Nal-Pro-NH2	100% ^c	100% ^c	100% ^c	0.006	
4	Tyr-Pro-Phe-Phe-NH ₂ (endomorphin-2) ^b	13.70 ± 2.30	24.80 ± 3.00	64.70 ± 8.00	1.83	
5	Tyr-Pro-d-1-Nal-Phe-NH2 ^b	$\textbf{6.88} \pm \textbf{1.21}$	15.60 ± 3.37	40.12 ± 6.06	8.32	
6	Dmt-Pro- _D -1-Nal-Phe-NH ₂	$\textbf{31.09} \pm \textbf{3.91}$	$\textbf{66.70} \pm \textbf{4.67}$	95.69 ± 2.27	0.85	

Table 3 – Antinociceptive effect of morphiceptin and endomorphin-2 analogs in the mouse hot plate test after i.c.v. injection (10 μ g/animal)

All values are expressed as mean \pm S.E.M. (n = 10).

^a Data from Ref. [6].

^b Data from Ref. [7].

 $^{\rm c}\,$ Percentage of mice with latency $\ge\!\!240\,s$ (cut-off time).



Fig. 3 – Antagonist effect of β -funaltrexamine (β -FNA, 1 µg) on the hot plate inhibition of paw licking (PL), rearing (R), and jumping (J) induced by i.c.v. injection of A) EM-2 (3 µg), B) [Dmt¹, D-1-Nal³]morphiceptin (analog 3, 0.01 µg), and C) [Dmt¹, D-1-Nal³]EM-2 (analog 6, 1 µg), determined in the hot plate assay. The data represent the mean \pm S.E.M. of 10 mice per group. A two-way ANOVA analysis revealed a

3. Results and discussion

3.1. Structure-activity relationship studies

In our previous studies it was demonstrated that the introduction of naphtylalanine residues into the sequence of various MOP receptor-selective peptide ligands was advantageous for their activity [5,6,15]. Here we report on the synthesis and pharmacological characterization of morphiceptin and EM-2 analogs with D-1-Nal substituted for Phe in position 3 and containing Dmt in place of Tyr¹ (Table 1). Binding affinities of the new analogs for the MOP and DOP receptors in comparison with the parent peptides are listed in Table 2. In the series of morphiceptin analogs, replacement of Phe³ by D-1-Nal led to 2 with a 40-fold enhanced μ -affinity. The same replacement of Phe³ in EM-2 resulted in an approximately 30-fold drop in MOP receptor binding affinity, indicating that the Phe³ residue is more critical in EM-2 than in morphiceptin. Introduction of Dmt in position 1 resulted in a drastic (about two orders of magnitude) increase in MOP and, to a lesser extent, in DOP receptor binding affinity. The effect of Dmt¹ replacement is not surprising, since it was expected from the results of other Dmt¹-containing opioid peptide analogs [2,18,19,20,23,24]. The in vitro opioid agonist potencies were determined using the isolated guinea-pig ileum (GPI) assay for MOP receptors and the mouse vas deferens (MVD) assay for DOP receptors (Table 2). The results obtained with these functional assays were in agreement with the receptor binding affinities measured in the binding assays. Morphiceptin exhibited 40-fold lower GPI potency than EM-2 ($IC_{50} = 318$ and 7.71 nM, respectively). D-1-Nal substitution in positions 3 of morphiceptin resulted in an enormous increase in the MOP receptor agonist potency, while in EM-2 that same substitution produced a drastic potency decrease ($IC_{50} = 9.57$ and 872 nM, respectively). Substitution of Dmt for Tyr¹ in the latter two analogs greatly increased potency in the GPI and MVD assays.

3.2. Antinociceptive activity studies

It is well known that even a small change in the hydrophobicity and spatial conformation of an opioid ligand may not only affect its receptor affinity [1,7,21,24], but also its biological efficacy as an antinociceptive agent *in vivo* [7,14,26]. In that regard, the methyl groups on the phenolic ring in Dmt undoubtedly play a role in strengthening

significant interaction between β -funaltrexamine and endomorphin-2: F(1,36) = 17.743; ^cp < 0.001 (for paw licking), F(1,36) = 22.957; ^cp < 0.001 (for rearing), F(1,36) = 23.959; ^cp < 0.001 (for jumping); between β funaltrexamine and [Dmt¹, D-1-Nal³]morphiceptin (analog 3): F(1,36) = 23.162; ^cp < 0.001 (for paw licking), F(1,36) = 22.811; ^cp < 0.001 (for rearing), F(1,36) = 15.709; ^cp < 0.001 (for jumping); between β -funaltrexamine and [Dmt¹, D-1-Nal³]EM-2 (analog 6): F(1,36) = 29.823; ^cp < 0.001 (for paw licking), F(1,36) = 22.326; ^cp < 0.001 (for rearing), F(1,36) = 25.147; ^cp < 0.001 (for jumping). receptor binding, presumably by engaging in additional hydrophobic interactions.

The antinociceptive activity of the analogs was determined using the hot plate test (supraspinally mediated analgesia) after intracerebroventricular (i.c.v.) administration, in comparison with the effect produced by morphiceptin and EM-2. The obtained results show that a similar analgesic effect was produced by morphiceptin and EM-2 (Table 3). Much stronger activities were observed with analogs 2, 3, and 6. For the two most potent analogs 3 and 6 dose-response curves were obtained (Fig. 1). In the case of compound 3, the antinociceptive effect was observed at a dose as low as 0.1 ng, and for 6 around 10 ng, whereas in the case of both parent compounds much higher doses were required (100 ng per animal). In a time-course study of the antinociceptive effect, compound 3, morphiceptin, and EM-2 showed maximal response within 10 min after i.c.v. injection. The duration of the effect was about 20 min for morphiceptin, 60 min for EM-2, and 120 min for 3 (Fig. 2). The MOP receptor selective antagonist β funaltrexamine (β-FNA) effectively reversed the analgesic effect of all tested compounds, indicating that their action was mediated by MOP receptor (Fig. 3).

4. Conclusions

In conclusion, the novel analog [Dmt¹, D-1-Nal³]morphiceptin showed very high binding affinity for the MOP receptor and was a very potent MOP receptor agonist in the GPI functional assay. The exceptionally strong analgesic effect of this analog and its greatly increased duration of action in comparison with EM-2 place [Dmt¹, D-1-Nal³]morphiceptin among the most potent MOP receptor ligands published so far. The study demonstrated that D-1-Nal might be useful as a Phe surrogate in the design of the opioid analogs with unique biological activity.

Acknowledgements

This work was supported by the grants from the Medical University of Lodz (nos. 502-11-460, 502-11-461, and 503-1099-1), a grant "Start" from the Foundation for Polish Science (to J.F.), a grant from the Centre National de la Recherche Scientifique (CNRS, France), and grants from the Canadian Institutes of Health Research (MOP-5655) and the NIH, USA (DA-004443 and DA-008924). The authors wish to thank Jozef Cieslak for his excellent technical assistance.

- [1] Ambo A, Murase H, Niizuma H, Ouchi H, Yamamoto Y, Sasaki Y. Dermorphin and deltorphin heptapeptide analogues: replacement of Phe residue by Dmp greatly improves opioid receptor affinity and selectivity. Bioorg Med Chem Lett 2002;12:879–81.
- [2] Balboni G, Guerrini R, Salvadori S, Bianchi C, Rizzi D, Bryant SD, et al. Evaluation of the Dmt-Tic pharmacophore: conversion of a potent δ-opioid receptor antagonist into a

potent δ agonist and ligands with mixed properties. J Med Chem 2002;45:713–20.

- [3] Cardillo G, Gentilucci L, Qasem AR, Sgarzi F, Spampinato S. Endomorphin-1 analogues containing beta-proline are muopioid receptor agonists and display enhanced enzymatic hydrolysis resistance. J Med Chem 2002;45:2571–8.
- [4] Chang K-J, Lillian A, Hazum E, Cuatrecasas P, Chang J-K. Morphiceptin (NH4-Tyr-Pro-Phe-Pro-CONH2): a potent and specific agonist for morphine (mu) receptors. Science 1981;121:75–7.
- [5] Fichna J, do-Rego J-C, Chung NN, Lemieux C, Schiller PW, Poels J, et al. Synthesis and characterization of potent and selective μ -opioid receptor antagonists, [Dmt¹, D-2-Nal⁴]endomorphin-1 (Antanal-1) and [Dmt¹, D-2-Nal⁴]endomorphin-2 (Antanal-2). J Med Chem 2007;50: 512–20.
- [6] Fichna J, do-Rego J-C, Costentin J, Chung NN, Schiller PW, Kosson P, et al. Opioid receptor binding and in vivo antinociceptive activity of position 3-substituted morphiceptin analogs. Biochem Biophys Res Commun 2004;320:531–6.
- [7] Fichna J, do-Rego J-C, Costentin J, Janecka A. Characterization of antinociceptive activity of novel endomorphin-2 and morphiceptin analogs modified in the third position. Biochem Pharmacol 2005;69:179–85.
- [8] Fichna J, Gach K, Piestrzeniewicz M, Burgeon E, Poels J, Vanden Broeck J, et al. Functional characterization of opioid receptor ligands by aequorin luminescence-based calcium assay. J Pharmacol Exp Ther 2006;317:1150–4.
- [9] Fichna J, Janecka A, Bailly L, Marsais F, Costentin J, do-Rego J-C. In vitro characterization of novel peptide inhibitors of endomorphin-degrading enzymes in the rat brain. Chem Biol Drug Des 2006;68:173–5.
- [10] Fichna J, Janecka A, Costentin J, do-Rego J-C. The endomorphin system and its evolving neurophysiological role. Pharmacol Rev 2007;59:88–123.
- [11] Fujita Y, Tsuda Y, Li T, Motoyama T, Takahashi M, Shimizu Y, et al. Development of potent bifunctional endomorphin-2 analogues with mixed mu-/delta-opioid agonist and delta-opioid antagonist properties. J Med Chem 2004;47:3591–9.
- [12] Hansen Jr DW, Stapelfeld A, Savege MA, Reichman M, Hammond DL, Haaseth RC, et al. Systemic analgesic activity and δ -opioid selectivity in [2,6-dimethyl-Tyr¹,D-Pen⁵]enkephalin. J Med Chem 1992;35:684–7.
- [13] Janecka A, Fichna J, Kruszynski R, Sasaki Y, Ambo A, Costentin J, et al. Synthesis and antinociceptive activity of cyclic endomorphin-2 and morphiceptin analogs. Biochem Pharmacol 2005;71:188–95.
- [14] Jinsmaa Y, Marczak E, Fujita Y, Shiotani K, Miyazaki A, Li T, et al. Potent in vivo antinociception and opioid receptor preference of the novel analogue [Dmt1]endomorphin-1. Pharmacol Biochem Behav 2006;84:252–8.
- [15] Kruszynski R, Fichna J, do-Rego J-C, Chung NN, Schiller PW, Kosson P, et al. Novel endomorphin-2 analogs with μ -opioid receptor antagonist activity. J Pept Res 2005;66: 125–31.
- [16] Kruszynski R, Fichna J, do-Rego J-C, Janecki T, Kosson P, Pakulska W, et al. Synthesis and biological activity of Nmethylated analogs of endomorphin-2. Bioorg Med Chem 2005;13:6713–7.
- [17] Lengyel I, Orosz G, Biyashev D, Kocsis L, Al-Khrasani M, Ronai A, et al. Side chain modifications change the binding and agonist properties of endomorphin 2. Biochem Biophys Res Commun 2002;290:153–61.
- [18] Li T, Shiotani K, Miyazaki A, Tsuda Y, Ambo A, Sasaki Y, et al. Bifunctional [2',6'-dimethyl-Ltyrosine1]endomorphin-2 analogues substituted at position 3 with alkylated phenylalanine derivatives yield potent

REFERENCES

mixed mu-agonist/delta-antagonist and dual muagonist/delta-agonist opioid ligands. J Med Chem 2007;50:2753–66.

- [19] Okada Y, Fujita Y, Motoyama T, Tsuda Y, Yokoi T, Li T, et al. Structural studies of [2',6'-dimethyl-Ltyrosine1]endomorphin-2 analogues: enhanced activity and cis orientation of the Dmt-Pro amide bond. Bioorg Med Chem 2003;11:1983–94.
- [20] Sasaki Y, Ambo A, Murase H, Hirabuki M, Ouchi H, Yamamoto Y. In: Shioiri T, editor. Peptide Science 2000. Osaka: The Japanese Peptide Society; 2001. p. 117–20.
- [21] Sasaki Y, Sasaki A, Niizuma H, Goto H, Ambo A. Endomorphin 2 analogues containing Dmp residue as an aromatic amino acid surrogate with high mu-opioid receptor affinity and selectivity. Bioorg Med Chem 2003;11:675–8.
- [22] Sasaki Y, Suto T, Ambo A, Ouchi H, Yamamoto Y. Biological properties of opioid peptides replacing Tyr at position 1 by 2,6-dimethyl-Tyr. Chem Pharm Bull (Tokyo) 1999;47:1506–9.
- [23] Schiller PW, Fundytus ME, Merovitz L, Weltrowska G, Nguyen TM, Lemieux C, et al. The opioid mu agonist/delta antagonist DIPP-NH(2)[Psi] produces a potent analgesic effect, no physical dependence, and less tolerance than morphine in rats. J Med Chem 1999;42:3520–6.

- [24] Schiller PW, Nguyen TM-D, Berezowska I, Dupuis S, Weltrowska G, Chung NN, et al. Synthesis and in vitro opioid activity profiles of DALDA analogues. Eur J Med Chem 2000;35:895–901.
- [25] Schiller PW, Weltrowska G, Schmidt R, Nguyen TM-D, Berezowska I, Lemieux C, et al. Four different types of opioid peptides with mixed μ agonist/δ antagonist properties. Analgesia 1995;1:703–6.
- [26] Szeto HH, Lovelace JL, Fridland G, Soong Y, Fasolo J, Wu D, et al. In vivo pharmacokinetics of selective μ-opioid peptide agonists. J Pharmacol Exp Ther 2001;298:57–61.
- [27] Tömböly C, Kövér KE, Péter A, Tourwé D, Biyashev D, Benyhe S, et al. Structure-activity study on the Phe side chain arrangement of endomorphins using conformationally constrained analogues. J Med Chem 2004;47:735–43.
- [28] Zadina JE, Hackler L, Ge L-J, Kastin AJ. A potent and selective endogenous agonist for the mu-opiate receptor. Nature 1997;386:499–502.
- [29] Zhao QY, Chen Q, Yang DJ, Feng Y, Long Y, Wang P, et al. Endomorphin 1[psi] and endomorphin 2[psi], endomorphins analogues containing a reduced (CH2NH) amide bond between Tyr1 and Pro2, display partial agonist potency but significant antinociception. Life Sci 2005;77:1155–65.