

# Ligustilide induces vasodilatation via inhibiting voltage dependent calcium channel and receptor-mediated $\text{Ca}^{2+}$ influx and release

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## Abstract

The purpose of the present study was to investigate the effect of ligustilide on vasodilatation in rat mesenteric artery and the mechanisms responsible for it. Isometric tension of rat mesenteric artery rings was recorded by a sensitive myograph system in vitro. The results showed that ligustilide at concentrations more than 10  $\mu\text{M}$  relaxed potassium chloride (KCl)-precontracted rat mesenteric artery in a concentration-dependent manner. The vasodilatation effect of ligustilide was not dependent on endothelium. Ligustilide rightwards shifted concentration–response curves induced by KCl, calcium chloride ( $\text{CaCl}_2$ ), noradrenaline (NA) or 5-hydroxytryptamine (5-HT) in a non-parallel manner. This suggests that the vasodilatation effects were most likely via voltage-dependent calcium channel (VDCC) and receptor-operated calcium channel (ROCC). Propranolol, glibenclamide, tetraethylammonium and barium chloride did not affect the vasodilatation induced by ligustilide, showing that  $\beta$ -adrenoceptor, ATP sensitive potassium channel, calcium-activated potassium channel and inwardly rectifying potassium channel were not involved in the vasodilatation. Ligustilide concentration-dependently inhibited the vasoconstriction induced by NA or  $\text{CaCl}_2$  in  $\text{Ca}^{2+}$ -free medium, indicating that the vasodilatation relates to inhibition of extracellular  $\text{Ca}^{2+}$  influx through VDCC and ROCC, and intracellular  $\text{Ca}^{2+}$  release from  $\text{Ca}^{2+}$  store. Since caffeine-induced contraction was inhibited by ligustilide, inhibition of intracellular  $\text{Ca}^{2+}$  released by ligustilide occurred via the ryanodine receptors. Our results suggest that ligustilide induces vasodilatation in rat mesenteric artery by inhibiting the VDCC and ROCC, and receptor-mediated  $\text{Ca}^{2+}$  influx and release.

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**Keywords:** Ligustilide; Vasodilatation; Rat mesenteric artery; Calcium

## 1. Introduction

A lot of traditional Chinese medicines (TCM) have been widely used in clinic in lots of Asian countries for the treatment of cardiovascular diseases. In more than 10,000 different kinds of TCM, *Ligusticum chuanxiong* is a frequently used one for cardiovascular diseases to promote the circulation of blood and removing stasis (Hou et al., 2004a; Tsai et al., 2002; Gong and Sucher, 1999). The combination of *L. chuanxiong* Hort (or *Ligusticum wallichii* Franch.) (LC) and *Angelica sinensis* (Oliv.)

Diels (AS) is called *Fo Shou San* (Chinese medicinal formulas) with a long history of use for treatment of atherosclerosis and hypertension (Han et al., 1995).

Many effective components have been extracted from *L. chuanxiong*, such as chuanxiongzine (Huang et al., 1998; Cui et al., 2003), organic acids (Hou et al., 2004b), phthalides (Naito et al., 1996), etc. Ligustilide, one of the phthalides, has been studied and demonstrated that it has antiasthmatic action (Tao et al., 1984), centrally acting muscle relaxant effect (Ozaki et al., 1989), effects on atria (Nakazawa et al., 1989), and effects on central noradrenergic and/or GABA(A) systems (Matsumoto et al., 1998).

In addition, the effects of ligustilide on vasculature have been noticed and studied. It is mentioned that ligustilide is a main effective component of *L. chuanxiong* as a medicine in treatment for vascular diseases. It is reported that ligustilide has a vasodilatation effect on rat abdominal aorta, in the form of inhibiting NA

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and  $\text{CaCl}_2$ -induced vasoconstriction (Liang et al., 2005). However, the mechanism of this effect remain unknown. The objective of the present study is to investigate the mechanism responsible for ligustilide-induced vasodilatation in rat mesenteric artery.

## 2. Materials and methods

### 2.1. Animals and reagents

Male Sprague-Dawley rats (weighing 250–300 g) were from Animal Center of Xi'an Jiaotong University (Xi'an, China). Ligustilide (Lig) was supplied from Prof. He Langchong (Xi'an Jiaotong University, China). Noradrenaline (NA), 5-hydroxytryptamine (5-HT), acetylcholine (ACh), Triton X-100, propranolol (Prop), glibenclamide (Glib), tetraethylammonium (TEA), caffeine and dimethyl sulphoxide (DMSO) were obtained from Sigma Aldrich (St. Louis, USA). All other reagents were analytical reagent (AR) grade. All substances were dissolved in distilled water except for ligustilide which was dissolved in DMSO and further dilutions were made in Krebs's solution.

### 2.2. In vitro pharmacology

The rats were anesthetized and sacrificed by decapitation. The superior mesenteric artery was removed gently and immersed in cold oxygenated Krebs's solution and dissected free of adhering

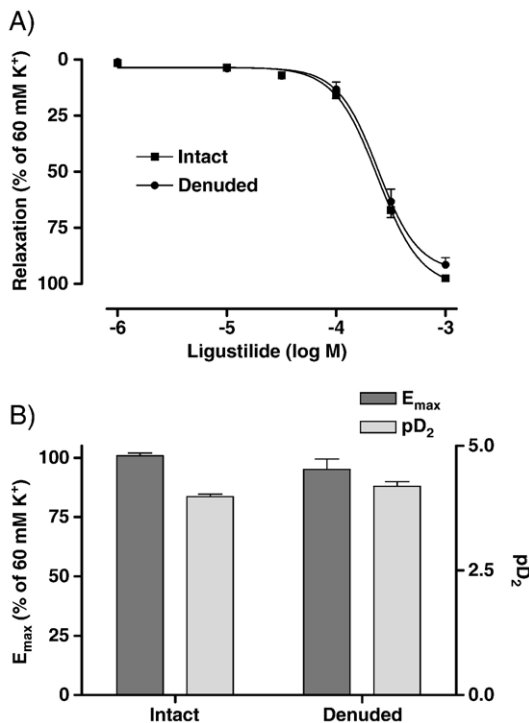


Fig. 1. (A) The concentration–response curves of ligustilide on vasodilatation effect of endothelium-intact and endothelium-denuded rat mesenteric artery precontracted by KCl. The relaxation was expressed as the percentage of the precontraction by 60 mM  $\text{K}^+$ . (B) Maximal relaxation and  $\text{pD}_2$  of ligustilide on rat mesenteric artery precontracted by KCl.  $R_{\text{max}}$  refers to maximal relaxation calculated as percentage of the corresponding precontraction with 60 mM  $\text{K}^+$ .  $\text{pD}_2$  is the negative logarithm of the drug concentration that elicited 50% relaxation.  $n=8$ .

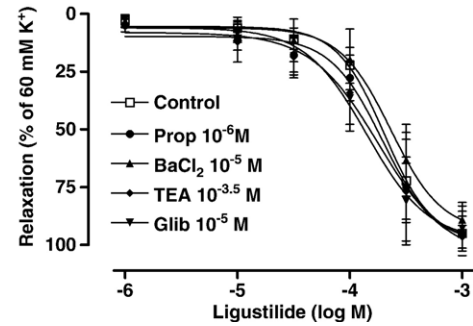


Fig. 2. Vasodilatation effect of ligustilide in endothelium-denuded artery rings in the presence of propranolol (Prop), glibenclamide (Glib), tetraethylammonium (TEA) or  $\text{BaCl}_2$ . No significant effects could be observed.  $n=7$ .

tissue under a microscope. In endothelium-denuded experiments the endothelium was denuded by perfusion of the vessel for 10 s with 0.1% Triton X-100 followed by another 10 s with Krebs's solution (Adner et al., 1996; Cao et al., 2004). The vessels were then cut into 1 mm long cylindrical segments and mounted on two L-shaped metal prongs, one of which was connected to a force displacement FT-03C transducer (Grass Instruments, USA) attached to a PowerLab 8SP unit (AD Instruments, UK) for continuous recording of the isometric tension, and the other to a displacement device. The mounted artery segments were immersed in tissue baths (DMT, Denmark) containing Krebs's solution of 1 ml, which was aerated continuously with a gas

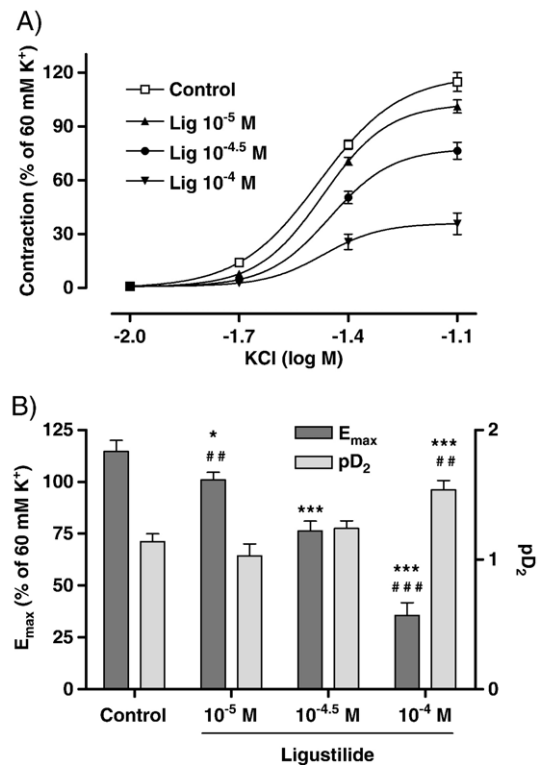


Fig. 3. (A) Effect of ligustilide on the concentration–response curves of KCl on rat mesenteric artery segments without endothelium. The concentration–response curve shifted towards right in a non-parallel manner compared with control. (B) Maximal effect ( $E_{\text{max}}$ ) and  $\text{pD}_2$  of KCl on rat mesenteric artery.  $n=8$ . \* $P<0.05$ , \*\*\* $P<0.001$  vs. control. ## $P<0.01$ , ### $P<0.001$  vs. ligustilide  $10^{-4.5}$  M.

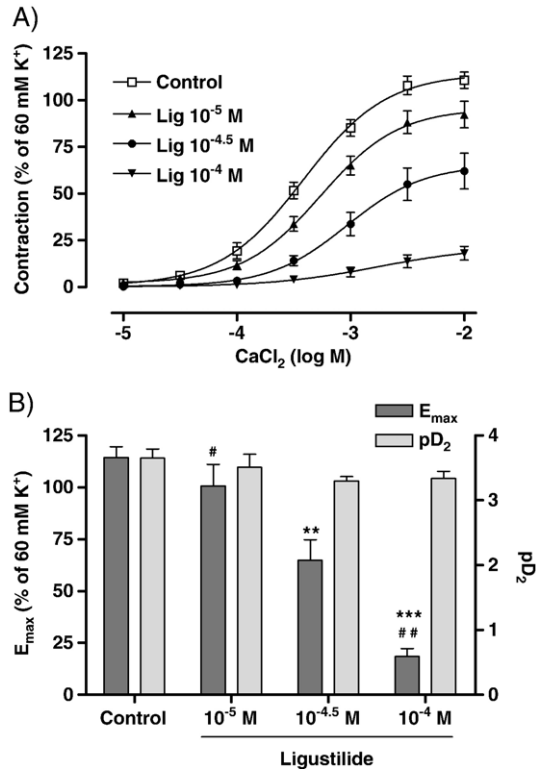


Fig. 4. (A) Effect of ligustilide on the concentration–response curve of  $\text{CaCl}_2$  on rat mesenteric artery segments without endothelium. The concentration–response curve shifted towards right in a non-parallel manner compared with control. (B)  $E_{\max}$  and  $pD_2$  of  $\text{CaCl}_2$  on rat mesenteric artery.  $n=7$ .  $**P < 0.01$ ,  $***P < 0.001$  vs. control.  $^{\#}P < 0.05$ ,  $^{\#\#}P < 0.01$  vs. ligustilide  $10^{-4.5}$  M.

mixture of 95%  $\text{O}_2$  and 5%  $\text{CO}_2$  and maintained at  $37^\circ\text{C}$ . The artery segments were equilibrated for 1.5 h with a resting tension of 5 mN before the experiments were started. The contractile capacity of each vessel segment was tested by exposure to a  $\text{K}^+$ -rich Krebs's solution (with 60 mM KCl) in which NaCl was exchanged for an equimolar concentration of KCl. When two reproducible contractions had been achieved the vessels were used for further experiments. After equilibration, artery segments were exposed to  $\text{K}^+$ -rich Krebs's solution. Once the sustained tension was obtained, ligustilide (1  $\mu\text{M}$ –1 mM) was added cumulatively to the baths, and the concentration–response curves to ligustilide were constructed.

In the experiment involving endothelium, the completeness of endothelium denudation was tested with acetylcholine (ACh) (10  $\mu\text{M}$ ) after pre-contraction with KCl. No relaxation in response to ACh in the denuded preparation indicated an effective functional removal of the endothelium. The rings with endothelium that produced less than 30% relaxation in response to ACh were discarded.

### 2.3. Statistical analysis

The effects of ligustilide are expressed as percentage of relaxation from the pre-contraction. Data are shown as mean  $\pm$  SEM. Statistical analysis was performed with unpaired Student's *t*-test. The *P* value of less than 0.05 was regarded to be significant.

## 3. Results

### 3.1. Vasodilation effect of ligustilide on rat mesenteric artery precontracted by KCl

Ligustilide (1  $\mu\text{M}$ –1 mM) concentration-dependently relaxed the artery segments pre-contracted by KCl with endothelium or without endothelium (Fig. 1A). In the artery ring segments with endothelium the maximum relaxation effect ( $R_{\max}$ ) of ligustilide was  $100.8 \pm 1.1\%$ . In rings denuded endothelium, the effect of ligustilide did not change significantly, and  $R_{\max}$  was  $95.0 \pm 4.4\%$  ( $P > 0.05$ , Fig. 1B).

### 3.2. Vasodilation effect of ligustilide in the presence of different blockers

The endothelium-denuded artery rings were incubated in the presence of propranolol (1  $\mu\text{M}$ ), glibenclamide (10  $\mu\text{M}$ ), tetraethylammonium (300  $\mu\text{M}$ ) or barium chloride ( $\text{BaCl}_2$ , 10  $\mu\text{M}$ ) for 20 min. Vasodilation effect of ligustilide on KCl-pre-contracted artery rings was recorded in order to test the effects of  $\beta$ -adrenoceptor, ATP sensitive potassium channel, calcium-activated potassium channel and inwardly rectifying potassium channel, which may contribute to the ligustilide-induced vasodilatation. The results (Fig. 2) showed that endothelium-denuded artery rings with propranolol, glibenclamide, tetraethylammonium or  $\text{BaCl}_2$  had no significant effect on the ligustilide-induced relaxation response.

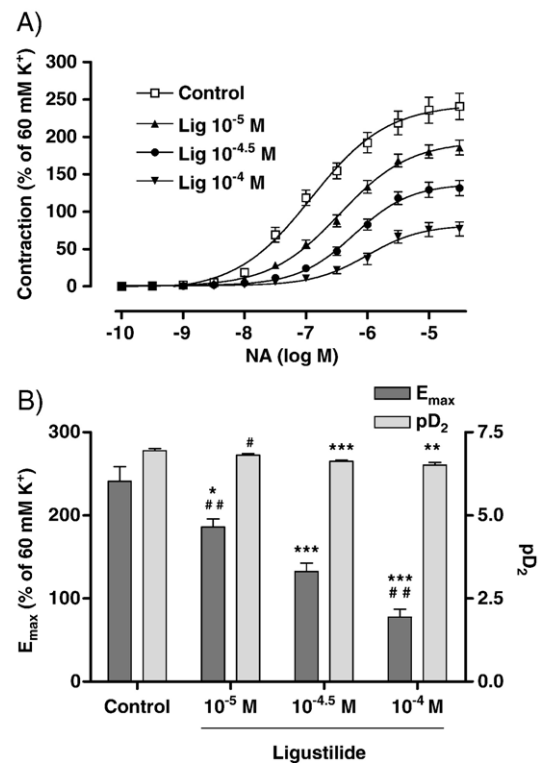


Fig. 5. (A) Effect of ligustilide on the concentration–response curve of NA on rat mesenteric artery segments without endothelium. The concentration–response curve shifted towards right in a non-parallel manner compared with control. (B)  $E_{\max}$  and  $pD_2$  of NA on rat mesenteric artery.  $n=7$ .  $*P < 0.05$ ,  $**P < 0.01$ ,  $***P < 0.001$  vs. control.  $^{\#}P < 0.05$ ,  $^{\#\#}P < 0.01$  vs. ligustilide  $10^{-4.5}$  M.

### 3.3. Effect of ligustilide on the KCl and CaCl<sub>2</sub>-induced concentration–contraction curves

After removal of endothelium, artery segments were equilibrated for 1.5 h. Krebs's solutions contained KCl (10, 20, 40, 80 mM) and DMSO of the same volume as ligustilide were replaced in order and the concentration–response curves were constructed. After washout and equilibrated for 1 h, the arteries were incubated with ligustilide (10, 30, 100 μM) for 15 min. Then, the concentration–response curves of KCl were constructed again as above, while DMSO was replaced with ligustilide. The concentration–response curve of KCl in the presence of ligustilide shifted towards right in a non-parallel manner compared with control ( $P < 0.05$ , Fig. 3A). The maximum effects ( $E_{\max}$ ) of KCl were reduced by different concentrations of ligustilide ( $P < 0.05$ , Fig. 3B). The  $pD_2$  value of ligustilide to KCl was  $4.28 \pm 0.05$ .

After contraction, the endothelium-denuded artery segments were exposed to Ca<sup>2+</sup>-free K<sup>+</sup>-rich solution containing EDTA (100 μM) and KCl (60 mM) for 20 min. After DMSO of the same volume as ligustilide was added to the baths and incubated for 15 min, CaCl<sub>2</sub> (10 μM to 10 mM) was added cumulatively to the baths, and the concentration–response curves to CaCl<sub>2</sub> were constructed. After washout and equilibration for 1 h, DMSO was replaced with ligustilide (10, 30, 100 μM) and the concentration–response curve of CaCl<sub>2</sub> were constructed again as above. The concentration–response curve of CaCl<sub>2</sub> in the presence of ligustilide shifted towards right in a non-parallel manner com-

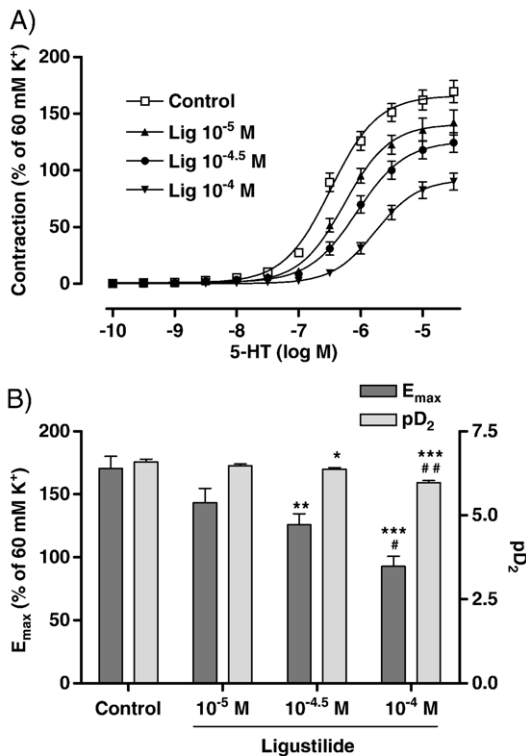


Fig. 6. (A) Effect of ligustilide on the concentration–response curve of 5-HT on rat mesenteric artery segments without endothelium. The concentration–response curve shifted towards right in a non-parallel manner compared with control. (B)  $E_{\max}$  and  $pD_2$  of 5-HT on rat mesenteric artery.  $n = 7$ . \* $P < 0.05$ , \*\* $P < 0.01$ , \*\*\* $P < 0.001$  vs. control. # $P < 0.05$ , ## $P < 0.01$  vs. ligustilide  $10^{-4.5}$  M.

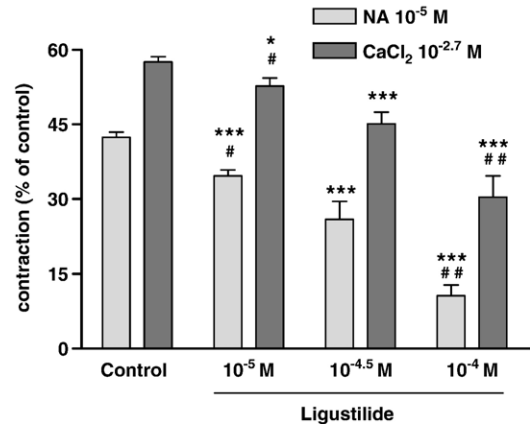


Fig. 7. Effect of ligustilide on the contraction induced by NA and CaCl<sub>2</sub> in Ca<sup>2+</sup>-free Krebs's solution.  $n = 8$ . \* $P < 0.05$ , \*\* $P < 0.001$  vs. control. # $P < 0.05$ , ## $P < 0.01$  vs. ligustilide  $10^{-4.5}$  M.

pared with control ( $P < 0.05$ , Fig. 4A). The  $E_{\max}$  of CaCl<sub>2</sub> were reduced by different concentrations of ligustilide ( $P < 0.05$ , Fig. 4B). The  $pD_2$  value of ligustilide to CaCl<sub>2</sub> was  $4.45 \pm 0.02$ .

### 3.4. Effect of ligustilide on the NA and 5-HT induced concentration–contraction curve

NA and 5-HT can both induce a potent and sustained constriction of mesenteric artery segments in a concentration-dependent manner. Ligustilide (10, 30, 100 μM) was added to the baths before cumulative addition of NA or 5-HT (0.1 nM–30 μM). As a result, ligustilide potently inhibited the NA or 5-HT induced vasoconstriction and concentration-dependently shifted the concentration–contractile curves towards right on a non-parallel manner with a decreased  $E_{\max}$  (Figs. 5 and 6). The  $pD_2$  value of ligustilide to NA and 5-HT was  $4.39 \pm 0.23$  and  $3.86 \pm 0.27$ , respectively.

### 3.5. Effect of ligustilide on the contraction of artery segments in Ca<sup>2+</sup>-free solution

After the contraction activity being tested, the endothelium-denuded artery segments were exposed to Ca<sup>2+</sup>-free Krebs's solution containing EDTA (100 μM) for 10 min. Another 10-min incubation of DMSO were followed by addition of 10 μM NA. After the NA-induced contraction being sustained, CaCl<sub>2</sub> of

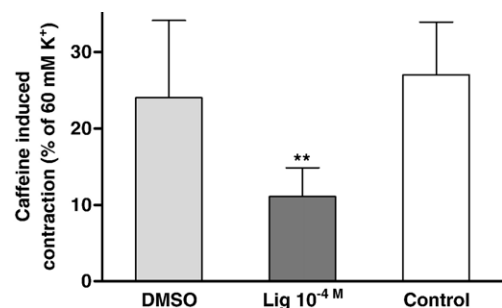


Fig. 8. Effect of ligustilide on the contraction induced by caffeine in Ca<sup>2+</sup>-free Krebs's solution. The contraction was expressed as the percentage of the precontraction by 60 mM K<sup>+</sup>.  $n = 7$ . \*\* $P < 0.01$  vs. control.

2 mM was added to contract the segments again (Broekaert and Godfriend, 1979; Zhao et al., 1997). The segments were thereafter washed with Krebs's solution (45 min contact time for  $\text{Ca}^{2+}$  refilling of the intracellular stores) and then 3 times with  $\text{Ca}^{2+}$ -free solution (15 min contact time). The second contractile response to NA and  $\text{CaCl}_2$  were tested in the presence of ligustilide (10, 30, 100  $\mu\text{M}$ ) for 10 min. Then the effect of ligustilide on the contraction induced by NA and  $\text{CaCl}_2$  in the  $\text{Ca}^{2+}$ -free Krebs's solution was tested in endothelium-denuded preparations. Ligustilide at three different concentrations significantly inhibited the contraction induced by NA and  $\text{CaCl}_2$  concentration-dependently ( $P < 0.05$ , Fig. 7).

### 3.6. Effect of ligustilide on the contraction induced by caffeine in $\text{Ca}^{2+}$ -free solution

The endothelium-denuded artery segments were exposed to  $\text{Ca}^{2+}$ -free Krebs's solution for 10 min followed by another 10-min incubation of DMSO. Then sustained contractions of arteries to caffeine (30 mM) were obtained (Cao et al., 2005). After washout and reloading, the effect of ligustilide (100  $\mu\text{M}$ ) on the vasoconstriction induced by caffeine was tested as above. A blank control (without incubation) was made before the end of the experiment (Fig. 8). Compared with control, the inhibitory rate in presence of DMSO was  $14.5 \pm 8.1\%$  ( $n = 7$ ), while that in presence of ligustilide was  $59.3 \pm 3.2\%$  ( $n = 7$ ,  $P < 0.01$ ). The results showed that ligustilide could significantly inhibit the vasoconstriction induced by caffeine in  $\text{Ca}^{2+}$ -free solution.

## 4. Discussion

The present study showed that ligustilide at concentration of more than 10  $\mu\text{M}$  relaxed the mesenteric artery precontracted by KCl. Removal of the endothelium did not affect the relaxant effect of ligustilide, indicating that ligustilide-induced vasodilatation was not dependent on endothelium.

Different blockers has been used to test the possible mechanisms. Propranolol (a general  $\beta$ -adrenoceptor antagonist), glibenclamide (an ATP sensitive potassium channel inhibitor), tetraethylammonium (a calcium-activated potassium channel inhibitor) and  $\text{BaCl}_2$  (an inwardly rectifying potassium channel inhibitor) did not affect the vasodilation induced by ligustilide, showing that  $\beta$ -adrenoceptor, ATP sensitive potassium channel, calcium-activated potassium channel and inwardly rectifying potassium channel were not involved in the vasodilatation.

The mechanism of vascular smooth muscle contraction involves different signal transduction pathways, all of which converge to increase intracellular calcium (Broekaert and Godfriend, 1979). Both extracellular  $\text{Ca}^{2+}$  influx, through voltage-dependent calcium channel (VDCC) or receptor-operated calcium channel (ROCC), and intracellular  $\text{Ca}^{2+}$  release result in the increase of intracellular calcium level. Ligustilide shifted the concentration–response curve of KCl and  $\text{CaCl}_2$  towards right in a non-parallel manner with decreased  $E_{\text{max}}$ , suggesting that VDCC on smooth muscle cells accounts for the vasodilator action of ligustilide. Moreover, the concentration–response curve of NA and 5-HT also shifted in the same pattern in the presence of

ligustilide, suggesting that ROCC was involved in the vasodilation effect of ligustilide.

The release of intracellular stored  $\text{Ca}^{2+}$  is mainly regulated by  $\text{IP}_3$  receptor system ( $\text{IP}_3\text{Rs}$ ) and ryanodine receptor system (RyRs). The former induces  $\text{Ca}^{2+}$  release directly when the receptors are bound to  $\text{IP}_3$ . The later may function through a  $\text{Ca}^{2+}$  induced  $\text{Ca}^{2+}$  release (CICR) mechanism when the receptors are activated by caffeine (Leijten and Van Breemen, 1984). The present results show that ligustilide concentration-dependently inhibited the contraction induced by NA and  $\text{CaCl}_2$  in  $\text{Ca}^{2+}$ -free medium, which suggests that the mechanism of the vasodilatation is related to the inhibition of extracellular  $\text{Ca}^{2+}$  influx through VDCC and ROCC, and intracellular  $\text{Ca}^{2+}$  release from  $\text{Ca}^{2+}$  store. Besides, ligustilide affects the caffeine-induced contraction of artery segments, which proved the involvement of RyRs in the release of intracellular stored  $\text{Ca}^{2+}$ .

In conclusion, ligustilide induced vasodilatation mainly by inhibiting voltage-dependent calcium channel and the receptor-mediated  $\text{Ca}^{2+}$  influx and release.

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